# **PI5I4 Acquired Carbapenemase-encoding Genes Among Gram-negative Isolates in USA Medical Centers: Report from the MYSTIC Program (2007)** RE Mendes, PR Rhomberg, M Castanheira, LM Deshpande, RN Jones JMI Laboratories, North Liberty, IA, USA

## ABSTRACT

**Objective:** To evaluate the occurrence of acquired carbapenemase-encoding genes among Gram-negative patient isolates recovered from USA medical centers (MC) in the 2007 MYSTIC Program.

Methods: 1,392 Enterobacteriaceae (ENT), 133 Acinetobacter spp. (ASP) and 465 Pseudomonas spp. (PSP) clinical isolates from 15 USA MC were susceptibility tested by CLSI reference broth microdilution method. ENT and PSP isolates showing MIC values  $\geq 2 \text{ mg/L}$  and  $\geq 16 \text{ mg/L}$ , respectively, for imipenem and meropenem were screened for KPC- and metallo-B-lactamase (MBL)-encoding genes. ASP isolates showing MIC values  $\geq$  16 mg/L for imipenem and meropenem were screened for MBL- and OXA-encoding genes. Presence of ISAbal upstream of the  $bla_{OXA-51}$  and clonality were also investigated by PCR and PFGE, respectively. Additionally, ASP isolates from previous MYSTIC Program years were tested for OXA genes.

**Results:** 42 (3.0% of the total) ENT isolates were screened and 35 (2.5%) isolates from MC located in New York (24/35; 68.6%), New Jersey (10/35; 28.6%) and Ohio (1/35; 2.9%) harbored bla<sub>KPC</sub>. This gene was found in C. freundii (2), E. coli (1), E. cloacae (3), K. oxytoca (1) and K. pneumoniae (28). Among 133 ASP, 50 (37.6%) isolates were screened and *bla*<sub>OXA-23</sub> was detected in 10 isolates from MC in New York (3), Kentucky (3), Arkansas (2) and Colorado (2), while bla<sub>OXA-24</sub> was detected in 5 isolates from New Jersey. ISAbal was associated with bla<sub>OXA-51</sub> in 78.8% of the cases and clonal dissemination of  $bla_{OXA}$ —carrying ASP within MC was noted. In the retrospective sample of ASP collected from 1999 to 2006, 42 (6.8%) isolates were screened for  $bla_{OXA}$ .  $bla_{OXA-23}$  was observed in 2 isolates from Tennessee (2004) and one isolate each from Kentucky and Washington (2005). Among the 36 (7.7%) PSP isolates screened, no carbapenemases were detected. MBL-encoding genes were not observed.

**Conclusions:** Although the results presented here document an alarming presence of  $bla_{KPC}$ , the rate (2.5%) was lower when compared with the previous MYSTIC year (4.5%; P = 0.0063; OR = 1.80, CI 95% 1.15-2.82). However, the KPC gene has spread among several different ENT species, while it was detected only in Klebsiella spp. in the previous year.  $bla_{OXA}$  genes have been detected in several MYSTIC Program MC within the USA; first detected in 2004. bla<sub>OXA</sub> genes were only observed in consecutive years in Kentucky, suggesting a sporadic appearance in ASP, overall. MBL-encoding genes are still quite rare in the MYSTIC Program.

#### INTRODUCTION

B-lactam antimicrobial agents that once were highly active against most Gramnegative pathogens are now less effective due to emerging resistance mechanisms harbored by Enterobacteriaceae, Pseudomonas aeruginosa and Acinetobacter baumannii. Among B-lactam resistance mechanisms, B-lactamases are most worrisome due to their ability to rapidly disseminate, since genes encoding most B-lactamases reside on mobile plasmids or transposons.

Several B-lactamase groups are of special concern because of their epidemiologic impact or possible limitations on contemporary antimicrobial therapy. Acquired carbapenemases (i.e. metallo-B-lactamases [MBLs], oxacillinases and serinecarbapenemases) can have a significant adverse impact on the clinical utility of carbapenems, and carbapenemase-producing isolates are usually non-susceptible to the vast majority of B-lactam agents.

The main objective of this study was to evaluate the occurrence of acquired carbapenemase-encoding genes among Gram-negative isolates recovered from United States (USA) medical centers participating in the 2007 MYSTIC Program. Additionally, Acinetobacter spp. from previous MYSTIC Program years (1999-2006) were screened for acquired oxacillinases.

# MATERIALS AND METHODS



Bacterial isolates. A total of 1,392 Enterobacteriaceae, 133 Acinetobacter spp. and 465 Pseudomonas spp. clinical isolates were consecutively collected from patients in 15 medical centers geographically dispersed across the USA. These strains were recovered from hospitalized patients with each participating institution contributing 200 isolates including Gram-positive and -negative species. Only one isolate per patient from a documented infection was included in the MYSTIC Program 2007 study. Species identification was confirmed by standard biochemical tests and use of the Vitek System (bioMerieux, Hazelwood, MO), when necessary.

Antimicrobial susceptibility testing. All isolates were tested for antimicrobial susceptibility using the broth microdilution method as described by the Clinical and Laboratory Standards Institute (CLSI, formerly the NCCLS; M7-A7, 2006). Cationadjusted Mueller-Hinton broth was used in validated panels manufactured by TREK Diagnostics (Cleveland, OH). Categorical interpretations for all antimicrobials were those found in MI00-SI8 (CLSI, 2008). Quality control (QC) was performed using Escherichia coli ATCC 25922, Staphylococcus aureus ATCC 29213 and P. aeruginosa ATCC 27853. All QC results were within ranges as published in CLSI documents.

Genotypic detection of carbapenemase-encoding genes. Enterobacteriaceae and Pseudomonas spp. isolates showing MIC values  $\geq 2 \text{ mg/L}$  and  $\geq 16 \text{ mg/L}$ , respectively, for imipenem and meropenem were screened for KPC- and MBL-encoding genes. Generic primers were designed to amplify all KPC- (KPC-1 through -5) and MBL-(IMP-like, VIM-like, SPM-I, SIM-I) encoding genes available in the GenBank using a multiplex PCR approach. A similar approach was also used to amplify other serinecarbapenemases (IMI-like, NMC-A and SME-like) among KPC-negative isolates.

Acinetobacter spp. isolates showing MIC values  $\geq 16$  mg/L for imipenem and meropenem were screened for MBL- and OXA- (OXA-23-, OXA-24-, OXA-51- and OXA-58-clusters) encoding genes. Upstream DNA sequences of bla<sub>OXA-51</sub> were studied by PCR using primers targeting ISAba1, 2 or 3. Additionally, 42 Acinetobacter spp. isolates exhibiting MIC values  $\geq$ 16 mg/L for imipenem and meropenem from prior MYSTIC Program years (1999-2006) were tested for OXA genes.

Molecular typing and sequencing analysis. Acinetobacter spp. chromosomal DNA was subjected to pulsed-field gel electrophoresis (PFGE) after digestion with Smal. PCR amplicons were sequenced on both strands and the nucleotide sequences and deduced amino acid sequences were analyzed using the Lasergene software package (DNASTAR, Madison, WI). Sequences were compared to others available via internet sources (http://www.ncbi.nlm.nih.gov/blast/).

#### RESULTS

- A total of 42 (3.0% of the total) Enterobacteriaceae isolates were screened and 35 (2.5%) isolates from medical centers located in New York (24/35; 68.6%), New Jersey (10/35; 28.6%) and Ohio (1/35; 2.9%) harbored  $bla_{KPC-2}$  (Table and Figure I).
- bla<sub>KPC-2</sub> was also detected in Citrobacter freundii (2), E. coli (1), Enterobacter cloacae (3), Klebsiella oxytoca (1) and K. pneumoniae (28; Table I). Other serine-carbapenemases, such as IMI-like, NMC-A and SME-like were not identified among KPC-negative isolates.
- Among 133 Acinetobacter spp., 50 (37.6%) isolates were screened and bla<sub>OXA-23</sub> was detected in 10 isolates from medical centers in New York (3), Kentucky (3), Arkansas (2) and Colorado (2), while bla<sub>OXA-24</sub> was observed in 5 isolates from New Jersey (Table and Figure 1).

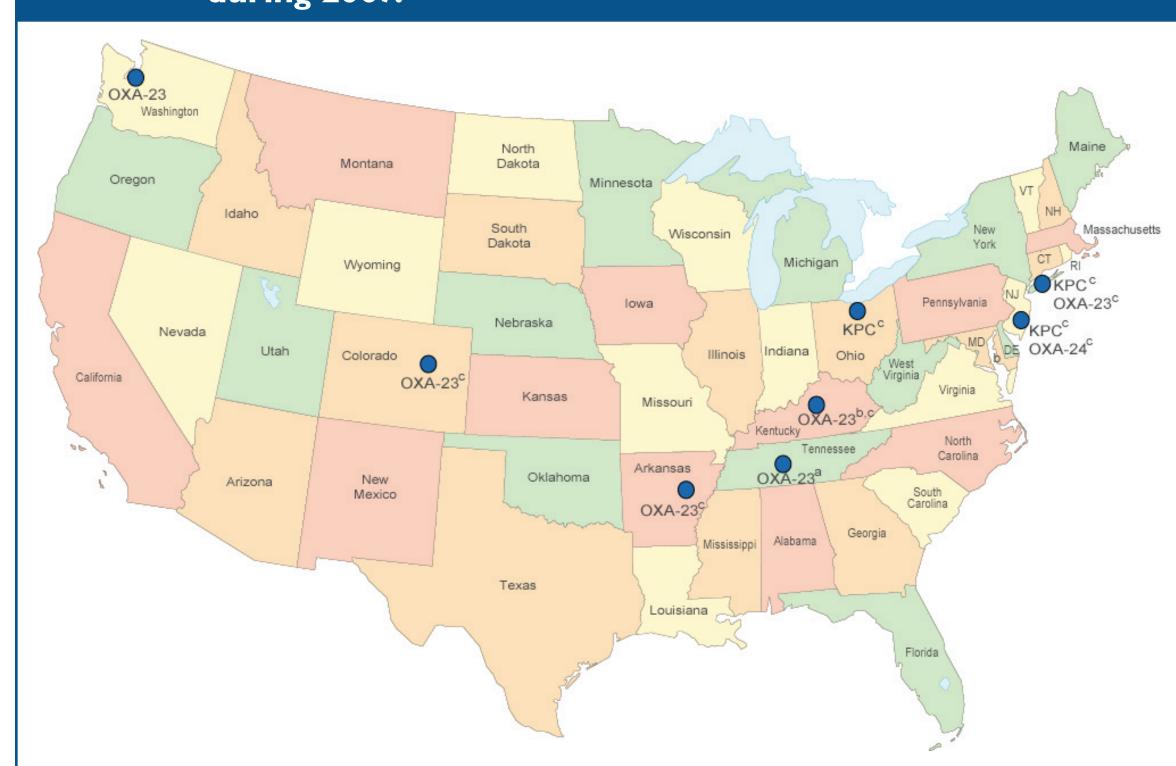


Contact details: JMI Laboratories North Liberty, IA, USA www.jmilabs.com 319.665.3370 319.665.3371 fax ronald-jones@jmilabs.com

- According to PFGE patterns, the three  $bla_{OXA-23}$ -carrying isolates from Kentucky were considered epidemiologically unrelated. The remaining OXA-23-producing Acinetobacter spp. were genetically related within specific medical institutions. The OXA-24-producing isolates recovered from a single hospital in New Jersey, belonged to a unique PFGE pattern (Table I).
  - Among Acinetobacter spp. showing negative results for acquired oxacillinases (OXA-24, -23 and -58), the ISAbal was detected upstream of the  $bla_{OXA-51}$  in 78.8%, which could explain the carbapenem-resistance phenotype.
  - A total of 42 (6.8%) Acinetobacter spp. isolates recovered between 1999 and 2006 were screened for  $bla_{OXA}$ .  $bla_{OXA-23}$  was observed in 2 isolates from Tennessee (2004) and one isolate each from Kentucky and Washington (2005), showing its recent escalation in the 2007 sample.
  - Among the 36 (7.7%) Pseudomonas spp. isolates screened, no carbapenemases were detected.

ß-lactamase (no. of isolates)	Medical center location (no. of isolates)	Bacterial species (no. of isolates)
KPC-2 (35)	New York, New York (15) New Brunswick, New Jersey (10) Mineola, New York (9) Cleveland, Ohio (1)	K. pneumoniae (28), Enterobacter cloacae (3), Citrobacter freundii (2), E. coli (1), K. oxytoca (1)
OXA-23 (I0)	Lexington, Kentucky (3) New York, New York (3) Denver, Colorado (2) Little Rock, Arkansas (2)	A. baumannii (10) <sup>a</sup>
OXA-24 (5)	New Brunswick, New Jersey (5)	A. baumannii (5) <sup>a</sup>

Figure I. Distribution of *bla*<sub>KPC-2</sub>, *bla*<sub>OXA-23</sub> and *bla*<sub>OXA-24</sub> detected in Enterobacteriaceae and A. baumannii isolates within the USA. <sup>a</sup> represents A. baumannii isolates recovered during 2004; <sup>b</sup> represents A. baumannii isolates recovered during 2005; and <sup>c</sup> represents A. baumannii isolates recovered during 2007.





## CONCLUSIONS

- Although these 2007 MYSTIC Program results presented here document an alarming presence of  $bla_{KPC-2}$ , the rate (2.5%) was significantly lower when compared with the previous MYSTIC Surveillance year (4.5%; P = 0.0063; OR = 1.80, CI 95% 1.15-2.82).
- Most KPC-producing isolates in this study were recovered from the New York City area, confirming the continued presence of this serious resistance problem.
- While the rate of *bla*<sub>KPC</sub>-carrying Enterobacteriaceae isolates was lower overall, this gene has spread among several different species, whereas it was detected only in Klebsiella spp. in earlier sampling.
- While  $bla_{OXA}$  genes have now been detected in Acinetobacter spp. collected from several MYSTIC Program medical centers within the USA (first detected in 2004), *bla<sub>OXA</sub>* genes were only observed in consecutive years in Kentucky, suggesting a rather sporadic appearance in Acinetobacter spp.; but recent expansion overall

#### ACKNOWLEDGEMENT

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### SELECTED REFERENCES

Clinical and Laboratory Standards Institute. (2006). M7-A7, Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard seventh edition. Wayne, PA: CLSI.

Clinical and Laboratory Standards Institute. (2008). M100-S18, Performance standards for antimicrobial susceptibility testing, 18th informational supplement. Wayne, PA: CLSI.

Mendes RE, Kiyota KA, Monteiro J, Castanheira M, Andrade SS, Gales AC, Pignatari AC, Tufik S (2007). Rapid detection and identification of metallo-B-lactamaseencoding genes by multiplex real-time PCR assay and melt curve analysis. J Clin Microbiol 45: 544-547.

Poirel L, Nordmann P (2006). Genetic structures at the origin of acquisition and expression of the carbapenem-hydrolyzing oxacillinase gene bla<sub>OXA-58</sub> in Acinetobacter baumannii. Antimicrob Agents Chemother 50: 1442-1448.

Poirel L, Pitout JD, Nordmann P (2007). Carbapenemases: Molecular diversity and clinical consequences. Future Microbiol 2: 501-512.

Woodford N, Ellington MJ, Coelho JM, Turton JF, Ward ME, Brown S, Amyes SG, Livermore DM (2006). Multiplex PCR for genes encoding prevalent OXA carbapenemases in Acinetobacter spp. Int J Antimicrob Agents 27: 351-353.