

ACTIVITY OF FAROPENEM AGAINST NEISSERIA GONORRHOEAE INCLUDING CONTEMPORARY ANTIMICROBIAL RESISTANCE PHENOTYPES

R.N. JONES, I.A. CRITCHLEY, N. JANJIC, S. POTTUMARTHY
JMI Laboratories, North Liberty, IA; Replidyne Inc., Louisville, CO

E-312

PATIENTS AND METHODS

STRAIN COLLECTION

Background: There is a worldwide concern about the escalating resistance (*R*) among *N. gonorrhoeae*, inclusive of 189 (71.3%) contemporary strains isolated from clinical specimens in 2004 enriched with 76 well-characterized resistant phenotypes of *N. gonorrhoeae* from the Jones Microbiology Institute (JMI); North Liberty, Iowa laboratories collection were analyzed.

The 189 recent clinical strains were obtained from six geographically distinct areas where QNG has been reported to be endemic/pedemic and include: California (7.3.7%), Hawaii (27.14.3%), New York (36.13.2%), Ohio (36.19.1%), Oregon (18.9.5%) and Washington State (76.40.2%). The 76 well-characterized resistant phenotypes included: 53 QNG isolates, 15 strains resistant to penicillin and tetracycline, and eight isolates with intermediate susceptibility to penicillin by mechanisms analysis. The majority of the QNG isolates (61.53.96.2%) were isolated in the Far East and characterized in Japan (courtesy of Professors Tanaka and Deguchi). More than half (31.60.8%) of these isolates have documented mutations in the quinolone-resistance-determining-regions (QRNG), with single aminoacid substitutions in GyA, 20 strains Ser-91 → Ile, 14 strains Asp-95 → Asn, 4 ParC, 8 strains GyA (Ser-91 → Phe), 1 ParC (Asp-86 → Ile), 6 strains) and GyA (Ser-91 → Phe), ParC (Ser-87 → Ile), 2 strains); and double aminoacid substitutions in GyA and single substitution in ParC, 3 strains (GyA (Ser-91 → Phe, Asp-95 → Asn), ParC (Ser-88 → Pro), 2 strains) and (GyA (Ser-91 → Phe, Asp-95 → Glu), ParC (Glu-91 → Glu), 1 strain).

Methods: 265 NG strains were analyzed: 189 recent genital tract strains and 76 well-characterized R phenotypes (53 QNG [31 from Japan, 1990s], 23 with reduced S to penicillin [PEN] and/or tetracycline [TET]). Agar dilution MICs were determined using GC agar base supplemented with IsoVitaleX or defined CLSI supplement. FAR activity was compared to: PEN, TET, ceftriaxone (CTX), cefuroxime (CIP), aztreonam (AZT), ciprofloxacin (CIP) and levofloxacin (LEV).

Results: FAR activity was adversely influenced by L-cysteine in IsoVitaleX (4X MIC₅₀) increasing 0.06 vs 0.05 mg/ml, R among NG to CIP, PEN and TET values > 24.2%. Beta order of potency was: CTX (MIC₅₀, 0.06 mg/ml) > FAR (0.25) > XM (1) > TET (2) > PEN = CIP = LEV (4). FAR was 4X more active than oral XM (MIC₅₀, 0.25 vs 1.0 mg/ml) but 4X less active than CTX (0.25 vs 0.06 mg/ml). Activity of FAR was not affected by penicillines PPNG, MIC₅₀ 0.12 mg/ml, but non-PPNG-PEN-R was affected by penicillines PPNG, MIC₅₀ 0.12 mg/ml. While the QNG rates for 6 states ranged in MIC₅₀ 0.016 mg/ml to 0.25 mg/ml PEN-R. While the QNG rates for 6 states ranged from 0.0% (Ohio) to 66.7% (Hawaii), FAR retained a potency (MIC₅₀, ≤ 0.25 mg/ml) against these strains and activity was maintained acceptable potency irrespective of the R phenotype methods. Ability of FAR to maintain acceptable potency irrespective of an oral β-lactam for uncomplicated NG infections where QNG has become prevalent.

Conclusions: FAR activity should be determined on L-cysteine-free agar by CLSI methods. Ability of FAR to maintain acceptable potency irrespective of the R phenotype makes it a promising candidate for therapeutic development as an oral β-lactam for uncomplicated NG infections where QNG has become prevalent.

RESULTS

Table 2. Faropenem activity compared to reference agents when tested against 265 isolates of *N. gonorrhoeae*

Antimicrobial agent	MIC (μg/ml)			% by category ^a
	Range	50%	90%	
Aztreonam	0.03 - 2	0.5	-	-
Ciprofloxacin	≤ 0.008 - > 4	0.008	4	60.4
Levofloxacin ^c	≤ 0.008 - > 4	0.016	4	61.9
Tetracycline	0.06 - > 4	0.5	2	20.4
Penicillin	0.016 - > 4	1	4	5.3
Ceftriaxone	≤ 0.008 - 0.12	≤ 0.008	0.06	100.0
Cefuroxime	≤ 0.008 - 0.12	≤ 0.008	0.06	100.0
Aztreomycin	-	-	-	-
Cefotaxime	-	-	-	-
Tetraacycline	0.06 - 2	0.5	2	86.3
Ciprofloxacin	0.06 - 2	0.25	1	0.0
Levobacitam ^c	0.12 - 4	0.25	1	0.0

a. Criteria for interpretation by the CLSI.
b. = no published CLSI interpretive criteria.
c. Breakpoint criteria of ciprofloxacin were used corrected for the 2-fold greater potency of levofloxacin.

b. = no published CLSI interpretive criteria.
c. Susceptible at ≤ 0.12 mg/ml; resistant at ≥ 1 mg/ml.

CONCLUSIONS

- Our results clearly confirmed that faropenem is effective *In vitro* against recent gonococcal isolates (MIC₅₀, 0.06 and MIC₉₀, 0.25 μg/ml). However, we observed that the activity of faropenem was adversely affected by L-cysteine or hydrochloride in the medium supplement IsoVitaleX (4-fold increase in MIC₅₀).
- Despite the preselection bias towards antimicrobial-resistant gonococcal isolates (entire collection QRNG rate of 39.6%; high-level penicillin and tetracycline resistance rates of 32.5% and 24.2%, respectively), the potency of faropenem was competitive, and was positioned between ceftriaxone and cefuroxime.
- While the activity of faropenem and the other β-lactams tested (ceftriaxone and cefuroxime) was not adversely affected by increasing oiprofloxacin MIC values, increasing penicillin MIC results were associated with an 8- to ≥ 32-fold increase in the MIC₅₀ value of faropenem and some comparators.
- In the analysis of recent clinical isolates collected in 2004 from six distinct areas in the USA, revealed that each faropenem MIC₅₀ value (0.06 μg/ml) was identical to that obtained for the entire collection (265 strains; Table 3).
- There was a marked impact of the geographical site on the endemic antimicrobial susceptibility rates for the isolates from all six states. Penicillin susceptibility rates varied from 0.0% (New York) to 52.8% (Ohio) (data not shown). Ciprofloxacin susceptibility rates varied markedly, ranging from 25.9% in Hawaii to greater than 90.0% in New York and Ohio (Table 3).
- The collection of 51 strains isolated from the Far East, with some being characterized in Japan (31 strains with documented QRNG mutations) were analyzed separately (Table 4). Faropenem retained excellent activity with MIC_{50/90} values of 0.12/0.25 μg/ml and with the highest MIC being only 0.25 μg/ml.
- Our results indicate that faropenem has excellent activity against this collection of gonococcal isolates analyzed, regardless of the resistance mechanism or phenotype. The potency of faropenem was comparable to tetracycline and penicillin were very high (≥ 28.0%). Despite these high rates of fluoroquinolone resistance, ceftriaxone, cefuroxime and faropenem remained active against these isolates (ceftriaxone 100.0% susceptible, faropenem MIC₅₀ ≤ 0.25 μg/ml).
- Our results indicate that faropenem has excellent activity against this collection of gonococcal isolates analyzed, regardless of the resistance mechanism or phenotype. The potency of faropenem was comparable to tetracycline and penicillin were very high (≥ 28.0%). Despite these high rates of fluoroquinolone resistance, ceftriaxone, cefuroxime and faropenem remained active against these isolates (ceftriaxone 100.0% susceptible, faropenem MIC₅₀ ≤ 0.25 μg/ml).

Table 1. Comparison of faropenem agar dilution MIC results obtained using the M7-A6 method and two growth supplements (with and without L-cysteine).

Supplement	MIC (μg/ml)			Cum. % inhibited at MIC (μg/ml)
	50%	90%	≤ 0.03	
A ^a	0.25	0.5	1.5	4.2 29.9 66.0 91.3 100.0
B ^b	0.06	0.25	26.0	58.1 88.3 99.6 100.0
Supplement	50%	90%	≤ 0.03	0.06 0.12 0.25 0.5 1
California (7)	0.016 - 0.25	0.06	-	97.2
Hawaii (27)	0.016 - 0.25	0.06	-	83.3
New York (25)	0.016 - 0.25	0.06	-	65.8
Ohio (36)	≤ 0.008 - 0.25	0.03	0.12	71.4
Oregon (18)	≤ 0.008 - 0.25	0.06	0.12	11.8
Washington (76)	≤ 0.008 - 0.25	0.06	0.12	80.4
All (189)	≤ 0.008 - 0.25	0.06	0.12	80.4
All strains (265)	≤ 0.008 - 0.5	0.06	0.25	80.4

a. A = supplement component of commercial IsoVitaleX containing high levels of L-cysteine.

b. B = 1% defined supplement recommended by the CLSI agar dilution method M7-A6 and M10-S15.

ACKNOWLEDGMENTS

The co-authors express their gratitude to the following contributors to this investigation: L. Deshpande, D.J. Bedenbach, M.G. Shwill and K.L. Meyer. We would also like to thank Joseph DiPietro, Summa Health Systems, Dwight Harry (University of Rochester Medical Center), Judy Lucco (PAES Regional Laboratory-Barberly and William L.P. Whittington (University of Washington) for their assistance in the collection of recent clinical isolates. This study was sponsored by an education/research grant by Replidyne, Inc.