C1024 In Vitro Activity of Minocycline Against Acinetobacter baumannii, Stenotrophomonas maltophilia and Burkholderia cepacia Isolated During 2013 from a Global Surveillance Program

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Abstract

Background

Minocycline (MIN) is a second generation tetracycline (TET) which is one of a few agents approved by the USA-FDA for treatment of Acinetobacter infections. In this study, the activity of MIN and comparators was evaluated against Acinetobacter baumannii-calcoaceticus species complex (AB), Stenotrophomonas maltophilia (SM), and Burkholderia cepacia species complex (BC) from patients at medical centers (2013).

Methods

AB, SM, and BC from 171 medical centers in 46 countries were susceptibility (S) tested against MIN and comparator agents. One isolate per infected patient episode was included. Local identifications were confirmed by the monitoring laboratory. S testing followed CLSI methods and quality control guidelines. CLSI and EUCAST interpretations were applied, where available. MDR were classified based on published recommendations using CLSI interpretive criteria (Magiorakos et. al.).

Results

MIN was among the most active agents tested against AB (72.3% S, $MIC_{50/90}$, 2/>8 µg/mL). S to MIN was 18.9, 23.1 and 24.3% greater than doxycycline (DOX) against all AB, MDR and XDR, respectively and 45.2, 54.2 and 55.8% greater than TET. MIN was the most active agent tested against SM (99.1% S, MIC₉₀, 2 µg/mL). After MIN, trimethoprimsulfamethoxazole (SMX, MIC_{50/90}, \leq 0.5/>4 µg/mL; 89.7% S) and levofloxacin (LEV, 74.0% S; MIC $_{50/90}, 1/\!\!>\!\!4~\mu\text{g/mL})$ were the next most active agents against SM. Against BC, SMX was the most active agent (MIC_{50/90}, ≤0.5/2 µg/mL; 100.0% S) followed by MIN (93.3% S, MIC₉₀, 4 µg/mL), ceftazidime (CAZ, 93.1% S; MIC₉₀, 4µg/mL), and meropenem (MEM, 89.7% S; MIC₉₀, 8 μg/mL).

Conclusions

MIN was among the most active agents against AB, and also exhibited a high level of activity for SM and BC. As an FDA-approved agent for treatment of infections due to *Acinetobacter* sp., MIN shows high activity in vitro, including MDR/XDR isolates. Clinical studies in infections due to SM and BC are warranted.

Organism (no. tested)	MIC _{50/90} in μg/mL (% S by CLSI):									
	MIN	DOX	TET	COL	LEV	MEM	CAZ	AMK	SMX	
Acinetobacter baumannii (1,312)	2/>8 (72.3)	2/>8 (53.4)	>32/>32 (27.1)	1/2 (96.4)	>4/>4 (19.8)	>8/>8 (23.5)	>32/>32 (19.7)	>32/>32 (31.5)	>4/>4 (29.5)	
MDR (1070)	4/>8 (66.2)	>8/>8 (43.1)	>32/>32 (12.0)	1/2 (95.9)	>4/>4 (2.2)	>8/>8 (6.4)	>32/>32 (3.0)	>32/>32 (16.3)	>4/>4 (14.1)	
XDR (943)	4/>8 (62.9)	>8/>8 (38.6)	>32/>32 (7.1)	1/2 (95.6)	>4/>4 (0.1)	>8/>8 (2.2)	>32/>32 (0.5%)	>32/>32 (12.3)	>4/>4 (9.2)	
Burkholderia <i>cepacia</i> (30)	2/4 (93.3)	4/8 ()	>32/>32 ()	>8/>8 ()	2/4 (86.2)	2/8 (89.7)-	2/4 (93.1%)	>32/>32 ()	≤0.5/2 (100.0)	
Stenotrophomo nas maltophila (464)	0.25/2 (99.1)	2/4 ()	16/32 ()	4/>8 ()	1/>4 (74.0)	>8/>8 ()	32/>32 (35.2)	>32/>32 ()	≤0.5/>4 (89.7)	

MDR = non-susceptible to ≥1 agent in ≥3 antimicrobial categories; XDR= non-susceptible to ≥1 agent in all but ≤2 antimicrobial classes; --, not available

Introduction

The tetracyclines were one of the early discovered broadspectrum antimicrobial classes which were found in the 1940s, with activity against Gram-positive and -negative bacteria. Development of enhanced compounds, second- and thirdgeneration semisynthetic compounds, has improved the range of coverage and/or improved oral bioavailability. One of the tetracyclines, minocycline, is among a few agents available which have been FDA-approved for treatment of infections due to Acinetobacter spp. The intravenous formulation of minocycline was recently granted status as a Qualified Infectious Disease Product (QIDP) by the US FDA for treatment of infections in patients with cystic fibrosis and chronic granulomatous disease (CGD) due to Stenotrophomonas maltophilia and Burkholderia cepacia.

Acinetobacter spp. is one of the ESKAPE pathogens for which there are limited treatment choices available. Acinetobacter spp. are frequently multidrug-resistant (MDR), so there are very few choices of antimicrobial treatments which are active against this bacterium. The usefulness of carbapenems against these organisms has diminished with the widespread occurrence of carbapenamase-mediated resistance. Polymyxins may be used, although there are questions about appropriate dosing, resistance development and toxicity.

This study evaluated the contemporary activity of minocycline tested against a collection of A. baumannii complex (hereafter referred to as A. baumannii), Stenotrophomonas maltophilia and *Burkholderia cepacia* species complex from patients at medical centers (2013) in North America, Europe and the Mediterranean region, Latin America and the Asia-Pacific.

Materials and Methods

Organisms: Isolates of *A. baumannii* (1,312), *S. maltophila* (464), and *B. cepacia* spp. complex (30) were selected from 171 medical centers in 46 countries including North America, Europe and the Mediterranean region (EU), Latin America, and the Asia Pacific. Only one isolate per infected patient episode was included and local organism identifications were confirmed by the monitoring laboratory (JMI Laboratories, North Liberty, Iowa, USA) using biochemical methods, the VITEK 2 System (bioMerieux, Hazelwood, Missouri, USA) and MALDI-TOF (Bruker Daltonics, Bellerica, Massachusetts, USA), as necessary.

Susceptibility Testing: Isolates were susceptibility tested using the reference broth microdilution method as described by the Clinical Laboratory and Standards Institute (CLSI). MDR and extensively drug-resistant (XDR) A. baumannii were classified as such per recently recommended guidelines (Magiorakos et. a; 2012), using the following antimicrobial class representative agents and CLSI non-susceptible interpretive criteria for A. baumannii: ceftazidime (\geq 16 µg/mL), meropenem (\geq 4 µg/mL), piperacillin/tazobactam (\geq 32/4 μ g/mL), levofloxacin (≥4 μ g/mL), gentamicin (≥8 μ g/mL), colistin (\geq 4 µg/mL), trimethoprim/sulfamethoxazole (\geq 4/76 μ g/mL) and tetracycline (\geq 8 μ g/mL). Classifications were based on the following recommended parameters: MDR = non-susceptible to \geq one agent in \geq three antimicrobial classes; XDR = non-susceptible to all but \leq two antimicrobial classes.

Results

Acinetobacter baumannii

A total of 81.6% of A. baumannii and were MDR (Table 1). The percentages of MDR varied by region from 61.7% in North America to 92.8% in EU (data not shown) XDR ranged from 51.2% in North America to 80.2% in Europe. Colistin was the most active agent tested against all A. baumannii with a MIC₉₀ value of 2 μ g/mL (96.4%) susceptible; Table 2). A. baumannii susceptibilities to other agents were less than 50% except for minocycline (MIC₅₀) value of 2 µg/mL; 72.3% susceptible) and doxycycline (53.4% susceptible). Figure 1 shows the cumulative MIC frequency distributions for all A. baumannii for minocycline and a number of comparative agents. Among the MDR A. baumannii, colistin was the most active agent exhibiting a MIC_{on} value of 2 µg/mL (95.9% susceptible: Table 2). Minocycline was the next most active agent exhibiting 66.2% susceptibility. Poorer susceptibility was noted with doxycycline (43.1%) and tetracycline (12.0%). Against XDR, colistin was the most active agent (MIC₉₀ 2 µg/mL; 95.6% susceptible; Table 2), followed by minocycline (62.9%) and doxycycline (38.6%). Susceptibility to other agents was less than 30% (Table 2).

Stenotrophomonas maltophilia and *Burkholderia cepacia* complex

There were only three antimicrobials that demonstrated potent activity against S. maltophilia (Table 3). Minocycline was the most active agent (MIC₉₀, 2 μ g/mL; 99.1% susceptible; Table 3). Doxycycline was slightly less active (MIC₉₀, 4 μ g/mL) and trimethoprim-sulfamethoxazole showed a MIC₉₀ of 4 µg/mL (89.7-91.7% [CLSI and EUCAST interpretive criteria]; Table 3). Minocycline activity ranged from 98.3% susceptible (Latin America) to 100.0% susceptible (Asia-Pacific) and trimethoprimsulfamethoxazole susceptibility was 87.2% (Asia-Pacific) to 92.4% (North America; data not shown).

There were only 30 Burkholderia cepacia complex isolates evaluated (Table 3). Trimethoprim-sulfamethoxazole was the most active agent (MIC₉₀, 2 μ g/mL; 100.0% susceptible; Table 3). Two other antimicrobials exhibited >90% susceptibility rates, and these were ceftazidime (MIC_{90} , 4 µg/mL; 93.1% susceptible) and minocycline (MIC₉₀, 4 μ g/mL; 93.3% susceptible; Table 3). Susceptibility to meropenem was 89.7% (MIC₉₀, 8 µg/mL; Table 3)

Table 1. Summary of minocycline activity tested against selected Gram-negative bacteria isolates from global surveillance (2013)

	No. of isolates (cumulative %) inhibited at minocycline MIC (μg/mL) of:									MIC (µg/mL)			
Organism (Number of isolates)	0.015	0.03	0.06	0.12	0.25	0.5	1	2	4	8	> 8	MIC ₅₀	MIC ₉₀
Acinetobacter baumannii complex (1,312)	6 (0.5)	40 (3.5)	100 (11.1)	109 (19.4)	126 (29.0)	160 (41.2)	109 (49.5)	120 (58.7)	178 (72.3)	199 (87.4)	165 (100.0)	2	> 8
MDR (1,070)	1 (0.1)	4 (0.5)	14 (1.8)	30 (4.6)	98 (13.7)	155 (28.2)	109 (38.4)	119 (49.5)	178 (66.2)	197 (84.6)	165 (100.0)	4	> 8
XDR (943)			8 (0.8)	12 (2.1)	68 (9.3)	130 (23.1)	100 (33.7)	105 (44.9)	170 (62.9)	191 (83.1)	159 (100.0)	4	> 8
Burkholderia cepacia species complex (30)						2 (6.7)	9 (36.7)	5 (53.3)	12 (93.3)	2 (100.0)		2	4
Stenotrophomonas maltophilia (464)		1 (0.2)	4 (1.1)	67 (15.5)	161 (50.2)	116 (75.2)	64 (89.0)	36 (96.8)	11 (99.1)	3 (99.8)	1 (100.0)	0.25	2

Table 3. Activity of minocycline and comparator agents

EUCAST^a

%S/%I/%R

- / - / -

- / - / -

-/-/-

- / - / -

-/-/-

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-/-/-

91.7 / 0.0 / 8.3

-/-/-

tested against Burkholderia cepacia complex and

Table 2. Activity of minocycline and comparator antimicrobial agents when tested against 1,312 isolates of Acinetobacter haumannii (all regions)

baumannii (all regions				Stenotrophomonas m			•	
Organism/ Antimicrobial agent		MIC ₉₀	CLSI ^a %S / %I / %R	EUCAST ^a %S / %I / %R	Organism/ Antimicrobial agent	MIC ₅₀	MIC ₉₀	CLSI ^a %S / %I / %R
Acinetobacter baumannii (1,312)					Burkholderia cepacia species complex	: (30)		
Minocycline	2	>8	72.3 / 15.1 / 12.6	-/-/-	Minocycline	2	4	93.3 / 6.7 / 0.0
Doxycycline	2	>8	53.4 / 0.9 / 45.7	-/-/-	-			
Tetracycline	>32	>32	27.1 / 10.8 / 62.1	- / - / -	Doxycycline	4	8	-/-/-
Piperacillin/tazobactam	>64	>64	16.1 / 3.7 / 80.2	- / - / -	Tetracycline	>32	>32	- / - / -
Ceftazidime	>32	>32	19.7 / 2.2 / 78.1	- / - / -	Dinaraoillin/tazahaatam		20	-/-/-
Meropenem	>8	>8	23.5 / 1.3 / 75.2	23.5 / 1.3 / 75.2	Piperacillin/tazobactam	4	32	-/-/-
Amikacin	>32	>32	31.5 / 4.0 / 64.5	29.5 / 1.9 / 68.6	Ceftazidime	2	4	93.1 / 6.9 / 0.0
Gentamicin	>8	>8	25.8 / 6.2 / 68.0	25.8 / 0.0 / 74.2	Marananam	2	8	89.7 / 6.9 / 3.4
Tobramycin	>16	>16	43.1 / 1.5 / 55.4	43.1 / 0.0 / 56.9	Meropenem	Ζ	0	09.7 / 0.9 / 3.4
Levofloxacin	>4	>4	19.8 / 5.3 / 74.9	19.3 / 0.5 / 80.2	Amikacin	>32	>32	- / - / -
Trimethoprim/sulfamethoxazole	>4	>4	29.5 / 0.0 / 70.5	29.5 / 3.1 / 67.4	Gentamicin	>8	>8	-/-/-
Colistin	1	2	96.4 / 0.0 / 3.6	96.4 / 0.0 / 3.6	Gentamicin	>0	>0	
MDR A. baumannii (1,070)					Tobramycin	>16	>16	- / - / -
Minocycline	4	>8	66.2 / 18.4 / 15.4	-/-/-	Levofloxacin	2	4	86.2 / 13.8 / 0.0
Doxycycline	>8	>8	43.1 / 1.1 / 55.8	-/-/-				
Tetracycline	>32	>32	12.0 / 12.6 / 75.4	- / - / -	Trimethoprim/sulfamethoxazole	≤0.5	2	100.0 / 0.0 / 0.0
Piperacillin/tazobactam	>64	>64	0.7 / 2.4 / 96.9	-/-/-	Colistin	>8	>8	-/-/-
Ceftazidime	>32	>32	3.0 / 1.8 / 95.2	-/-/-			20	- 1 - 1 -
Meropenem	>8	>8	6.4 / 1.7 / 91.9	6.4 / 1.7 / 91.9	Stenotrophomonas maltophilia (464)			
Amikacin	>32	>32	16.3 / 4.8 / 78.9	14.2 / 2.1 / 83.7	Minocycline	0.25	2	99.1 / 0.7 / 0.2
Gentamicin	>8	>8	10.0 / 7.5 / 82.5	10.0 / 0.0 / 90.0				
Tobramycin	>16	>16	30.9 / 1.8 / 67.3	30.9 / 0.0 / 69.1	Doxycycline	2	4	- / - / -
Levofloxacin	>4	>4	2.2/6.3/91.5	1.8/0.3/97.9	Tetracycline	16	32	-/-/-
Trimethoprim/sulfamethoxazole	>4	>4	14.1 / 0.0 / 85.9	14.1 / 3.6 / 82.3	· · · · · · · · · · · · · · · · · · ·			
Colistin	1	2	95.9 / 0.0 / 4.1	95.9 / 0.0 / 4.1	Piperacillin/tazobactam	>64	>64	- / - / -
XDR A. baumannii (943)					Ceftazidime	32	>32	35.2 / 13.3 / 51.5
Minocycline	4	>8	62.9 / 20.2 / 16.9	-/-/-				
Doxycycline	>8	>8	38.6 / 1.1 / 60.3	-/-/-	Meropenem	>8	>8	-/-/-
Tetracycline	>32	>32	7.1 / 12.7 / 80.2	-/-/-	Amikacin	>32	>32	- / - / -
Piperacillin/tazobactam	>64	>64	0.0/1.3/98.7	-/-/-				
Ceftazidime	>32	>32	0.5 / 1.4 / 98.1	-/-/-	Gentamicin	>8	>8	- / - / -
Meropenem	>8	>8	2.2/1.7/96.1	2.2/1.7/96.1	Tobramycin	>16	>16	- / - / -
Amikacin Gentamicin	>32	>32	12.3 / 5.1 / 82.6	10.4/1.9/87.7			. 1	740/405/455
	>8	>8	4.4 / 8.1 / 87.5	4.4/0.0/95.6	Levofloxacin	I	>4	74.0 / 10.5 / 15.5
Tobramycin	>16	>16	25.8/1.7/72.5	25.8/0.0/74.2	Trimethoprim/sulfamethoxazole	≤0.5	4	89.7 / 0.0 / 10.3
Levofloxacin	>4	>4	0.1/5.8/94.1	0.1/0.0/99.9	Colistin		. 0	-/-/-
Trimethoprim/sulfamethoxazole	>4	>4	9.2/0.0/90.8	9.2/3.6/87.2		4	>8	
Colistin		2	95.6 / 0.0 / 4.4	95.6 / 0.0 / 4.4	^a Criteria as published by the CLSI [20	151 and EU	CAST [201	[5], where available

Figure 1. Cumulative MIC frequency distribution for minocycline and comparator agents against all A. baumannii



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Conclusions

- Minocycline susceptibility rate against Acinetobacter baumannii (81.6% of them MDR) was 72.3%, the second highest after colistin (96.4% susceptible) and significantly higher than doxycycline (53.4%).
- Minocycline was the most potent agent against S. maltophilia (99.1% susceptible) closely followed by doxycycline and trimethoprim-sulfamethoxazole.
- Against the small collection of Burkholderia cepacia complex isolates (30), minocycline exhibited >93.3% susceptibility rate which was similar to ceftazidime; 100% of these strains were susceptible to trimethoprimsulfamethoxazole.
- Minocycline, particularly the intravenous formulation, has activity against several ESKAPE pathogens and merits consideration in seriously ill patients where treatment options may be limited due to the presence of MDR and XDR bacteria.

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References

Boucher HW, Talbot GH, Bradley JS, Edwards JE, Gilbert D, Rice LB, Scheld M, Spellberg B, Bartlett J (2009). Bad bugs, no drugs: no ESKAPE! An update from the Infectious Diseases Society of America. Clin Infect Dis 48: 1-12.

Chopra I, Roberts M (2001). Tetracycline antibiotics: Mode of action, applications, molecular biology, and epidemiology of bacterial resistance. Microbiol Mol Biol Rev 65: 232-260

Clinical and Laboratory Standards Institute (2015). M07-A10. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard tenth edition. Wayne, PA: CLSI.

Clinical and Laboratory Standards Institute (2015). M100-S25. Performance standards for antimicrobial susceptibility testing: 25th informational supplement. Wayne, PA: CLSI.

EUCAST (2015). Breakpoint tables for interpretation of MICs and zone diameters. Version 5.0, January 2015. Available at: http://www.eucast.org/clinical_breakpoints/. Accessed January 2015.

Farrell DJ, Flamm RK, Sader HS, Jones RN (2013). Antimicrobial activity of ceftolozane/tazobactam tested against Enterobacteriaceae and Pseudomonas aeruginos with various resistance patterns isolated in U.S. hospitals (2011-2012). Antimicrob Agents Chemother 57: 6305-6310

Grassi GG (1993). Tetracyclines - extending the atypical spectrum. Internat Journ Antimicrob Agents 3: S31-S46.

Jones RN, Guzman-Blanco M, Gales AC, Gallegos B, Lea Castro AL, Martino MD, Vega S, Zurita J, Cepparulo M, Castanheira M (2013). Susceptibility rates in Latin American nations: Report from a regional resistance surveillance program (2011). Braz J Infect Dis 17: 672-681.

Li H, Liu F, Zhang Y, Wang X, Zhao C, Chen H, Zhang F, Zhu B, Hu Y, Wang H (2015). Evolution of carbapenemresistant Acinetobacter baumannii revealed through wholegenome sequencing and comparative genomic analysis. Antimicrob Agents Chemother 59: 1168-1176

Li J, Rayner CR, Nation RL, Owen RJ, Spelman D, Tan KE, Liolios L (2006). Heteroresistance to colistin in multidrug-resistant Acinetobacter baumannii. Antimicrob Agents Chemother 50: 2946-2950.

Magiorakos AP. Srinivasan A. Carev RB. Carmeli Y Falagas ME, Giske CG, Harbarth S, Hindler JF, Kahlmeter G, Olsson-Liljeguist B, Paterson DL, Rice LB, Stelling J, Struelens MJ, Vatopoulos A, Weber JT Monnet DL (2012). Multidrug-resistant, extensively drug-resistant and pandrug-resistant bacteria: an international expert proposal for interim standard definitions for acquired resistance. Clin Microbiol Infect 18: 268-281.

Perez F, Hujer AM, Hujer KM, Decker BK, Rather PN, Bonomo RA (2007). Global challenge of multidrugresistant Acinetobacter baumannii. Antimicrob Agents Chemother 51: 3471-3484.

Talbot GH, Bradley J, Edwards JE, Jr., Gilbert D, Scheld M, Bartlett JG (2006). Bad bugs need drugs: an update on the development pipeline from the Antimicrobial Availability Task Force of the Infectious Diseases Society of America. Clin Infect Dis 42: 657-668.

Tuon FF, Rymsza AM, Penteado-Filho SR, Pilonetto M, Arend LN, Levin AS (2011). Should polymyxin be used empirically to treat infections in patients under high risk for carbapenem-resistant Acinetobacter? J Infect 62: 246-249.

Tygacil[®] Package Insert (2014). Available at www.tygacil.com. Accessed July 2015.

Yoon EJ, Courvalin P, Grillot-Courvalin C (2013). RND-type efflux pumps in multidrug-resistant clinical isolates of Acinetobacter baumannii: major role for AdeABC overexpression and AdeRS mutations. Antimicrob Agents Chemother 57: 2989-2995.