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Organism group/

Tetracycline

Daptomycin

Tigecycline<sup>c</sup>

Vancomycin

Trimethoprim/sulfamethoxazole

≤0.5 – 2

0.12 - 1

≤0.03 – 0.5

0.25 - 2

100.0 / -

0.5

0.25

0.12

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### **Abstract**

Background: Fusidic acid (FA) is an established antistaphylococcal agent used in clinical practice in Europe, Australia and Canada for at least three decades. FA is currently under clinical development for therapy of acute bacterial skin and skin-structure infections (ABSSSI) in the USA. This study assessed the activities of FA and comparators tested against *S. aureus* isolates.

Methods: S. aureus (7,340) were collected from 51 institutions distributed within all USA census regions in 2008 - 2009. Identification was performed by standard algorithms and Vitek 2. Isolates were tested for susceptibility (S) by CLSI methods (M07-A8 and M100-S20). S. aureus were analyzed based on resistance (R) patterns. A pan-R pattern was defined as S. aureus exhibiting a R phenotype to at least 5 antimicrobial classes (projected breakpoint).

Results: Isolates were mainly from bacteremia (46.0%), SSSI (31.5%) and respiratory tract infections (16.6%). Overall, FA inhibited 99.6% of tested S. aureus at ≤1 µg/mL FA (MIC<sub>50/90</sub>, 0.12/0.25  $\mu$ g/mL) and tigecycline (TG; MIC<sub>50/90</sub> 0.12/0.25 µg/mL; 100.0% S) showed equivalent activity against S. aureus, while FA was two- to 16-fold more active than daptomycin (DA; MIC<sub>50/90</sub>, 0.25/0.5 µg/mL; 99.9% S) vancomycin (VA; MIC<sub>50/90</sub>, 1/1  $\mu$ g/mL; 100.0% S) and linezolid (LZ; MIC<sub>50/90</sub>, 2/2 μg/mL; 99.9% S). Gentamicin (97.9% S), tetracycline (95.5% S) and trimethoprim/ sulfamethoxazole (98.6% S) also exhibited coverage against nearly all S. aureus. FA had consistent modal MIC and MIC<sub>50</sub> values (0.12 μg/mL) across all R subsets. Only FA (MIC<sub>oo</sub>,  $0.25 \mu g/mL$ ), TG (MIC<sub>90</sub>,  $0.25 \mu g/mL$ ), DA (MIC<sub>90</sub>, 0.5 $\mu$ g/mL), VA (MIC<sub>90</sub>, 1  $\mu$ g/mL) and LZ (MIC<sub>90</sub>, 2  $\mu$ g/mL) sustained potency against strains with pan-R patterns.

Conclusion: FA demonstrated potent activity against this current collection of *S. aureus* from USA hospitals. FA activity was comparable to TG, which were at least two-fold more active than other agents with similar clinical indications.

Organism/ Resistance pattern <sup>a</sup>	Number (cumulative %) inhibited at MIC (μg/mL)								
(no. tested/% of total)	≤0.06	0.12	0.25	0.5	1	2	4	8	
All S. aureus (7,340/100.0)	1070(14.6)	5327(87.2)b	826(98.4)	70(99.4)	20(99.6)	10(99.8)	9(99.9)	8(100.0	
2008 S. aureus (3,962/54.0)	397(10.0)	3066(87.4)	434(98.4)	48(99.6)	5(99.7)	3(99.8)	3(99.8)	6(100.0	
2009 S. aureus (3,378/46.0)	673(19.9)	2261(86.9)	392(98.5)	22(99.1)	15(99.6)	7(99.8)	6(99.9)	2(100.0	
MRSA (3,877/52.8)	493(12.7)	2884(87.1)	435(98.3)	38(99.3)	15(99.7)	8(99.9)	3(>99.9)	1(100.0	
MSSA (3,463/47.2)	577(16.7)	2443(87.2)	<u>391(98.5)</u>	32(99.4)	5(99.6)	2(99.6)	6(99.8)	7(100.0	
OX, ER, CP (1,364/18.6)	149(10.9)	1017(85.5)	184(99.0)	9(99.6)	3(99.9)	2(100.0)	_	_	
OX, ER, CL, CP (1,156/15.7)	97(8.4)	859(82.7)	168(97.2)	20(99.0)	7(99.6)	4(99.9)	1(100.0)	_	
OX, ER (751/10.2)	141(18.8)	<u>574(95.2)</u>	27(98.8)	3(99.2)	4(99.7)	0(99.7)	2(100.0)	_	
ER (729/10.0)	115(15.8)	523(87.5)	83(98.9)	5(99.6)	1(99.7)	1(99.9)	1(99.9)	1(100.0	
ER, CP (177/2.4)	27(15.3)	125(85.9)	20(97.2)	2(98.3)	2(99.4)	1(100.0)	_	_	
OX, CL, CP (137/1.9)	23(16.8)	104(92.7)	7(97.8)	2(99.3)	0(99.3)	0(99.3)	0(99.3)	1(100.0	
ER, CL, CP (112/1.5)	12(10.7)	83(84.8)	<u>16(99.1)</u>	1(100.0)	_	_	_	_	
Pan-R (170/2.4)	31(18.2)	116(86.5)	22(99.4)	22(99.0)	0(99.0)	1(100.0)	_	_	

methicillin-susceptible S. aureus. CL, clindamycin; CP, ciprofloxacin; ER, erythromycin; and OX, oxacillin. Pan-R = pan resistance phenotype (resistance to at least 5 antimicrobial classes). Criteria for susceptibility were those published by CLSI (M100-S20, 2010). Intermediate and resistant results grouped as resistant. Modal MIC and MIC<sub>50</sub> values are bold, while MIC<sub>90</sub> results are underlined.

### Introduction

Fusidic acid was first isolated in 1962 from Fusidium coccineum. This antimicrobial agent interacts with elongation factor G (EF-G), preventing its release from the ribosome and thereby inhibiting bacterial protein synthesis. Resistance to fusidic acid is primarily considered to be caused by mutations on the EF-G-encoding gene; however, acquired fusidic acid resistance mechanisms have also been described in clinical strains. These mobile genes, named fusB and fusC, can be chromosomal- or plasmid-mediated and were shown to protect EF-G from binding with fusidic acid molecules.

Fusidic acid has been used in Europe and Australia since the 1960's and in Canada since 1980; however this compound has <u>not</u> been licensed by the United States (USA) Food and Drug Administration (FDA); and it is not currently available for prescription in the USA. This belated introduction of fusidic acid into this country may now be viewed as positive in that it provides an additional antistaphylococcal agent with low toxicity and a unique mechanism of action that is devoid of cross resistance to other classes of antibacterials (including methicillin-resistant Staphylococcus aureus [MRSA]). Furthermore, the extensive foreign experience with fusidic acid in the treatment of serious staphylococcal infections over the past four decades provides a wealth of information about optimal use, particularly with regards to the implementation of dosing/delivery strategies to delay or avoid the development of resistance.

In this study, we evaluated the activity of fusidic acid against S. aureus isolates collected in USA medical centers during 2008 and 2009. Isolates were categorized according to different resistance patterns and mechanisms of resistance were evaluated for those strains showing elevated fusidic acid MIC results (≥2 µg/mL).

# Materials and Methods

Bacterial strains. A total of 7,340 S. aureus strains collected during 2008 and 2009 in 51 USA hospitals, located in the nine Census Regions were analyzed as part of the SENTRY Antimicrobial Surveillance Program. These isolates were collected from bloodstream, respiratory tract, and skin and skin-structure infections (SSSI), according to defined protocols. Only one isolate per patient from documented infections were included. Species identification was confirmed by standard biochemical tests, the Vitek System (bioMerieux, Hazelwood, MO) or 16S rRNA sequencing, when necessary.

Antimicrobial susceptibility testing. Isolates were susceptibility tested by a reference broth microdilution procedure as described by the Clinical and Laboratory Standards Institute (CLSI; 2009) using validated microdilution panels manufactured by TREK Diagnostics (Cleveland, OH, USA). Categorical interpretations for all antimicrobials were those found in M100-S20-U and quality control (QC) was performed using Escherichia coli ATCC 25922, S. aureus ATCC 29213 and Enterococcus faecalis ATCC 29212. All QC results were within specified ranges as published in CLSI documents. For fusidic acid, the interpretive susceptibility criteria of the EUCAST group (2010) were applied at ≤1 μg/mL.

Detection of fusidic acid resistance mechanisms. All strains displaying fusidic acid MIC at ≥2 µg/mL were tested for the presence of acquired fusB, fusC and fusD in a multiplex PCR approach. Detection of fusD (intrinsic of S. saprophyticus) was included in this reaction to detect strains that were incorrectly identified as other staphylococcal

Constitutive genes fusA and fusE were amplified and sequenced using Extensor Hi-fidelity Master Mix (ABGene, Sussex, United Kingdom) as well as custom and previously described oligonucleotides. Sequencing was performed in five and two reactions, respectively. The nucleotide sequences and deduced amino acid sequences were analyzed using the Lasergene software package (DNASTAR, Madison, WI) and compared with sequences available through the internet using BLAST (http://www.ncbi.nlm.nih.gov/blast/)

## Results

- Among 7,340 S. aureus collected in USA medical centers, 46.0% were recovered from bloodstream infections, 31.5% from SSSI and 16.6% from respiratory tract infections.
- Fusidic acid inhibited 99.6% of the S. aureus at ≤1 µg/mL (EUCAST susceptibility breakpoint). Twenty-seven (0.4%) strains showed MIC values at ≥2 µg/mL
- Fusidic acid was comparably active against *S. aureus* strains categorized into eight different resistance groups, including isolates resistant to five or more compounds (pan-resistant [pan-R] e.g., MIC<sub>50</sub> at 0.12 μg/mL for all eight groups, Table 1). The most common groups included resistances to oxacillin, erythromycin, clindamycin, and the fluoroquinolones.
- Among the orally administered agents evaluated (linezolid, tetracycline and trimethoprim/sulfamethoxazole), fusidic acid (modal MIC<sub>50</sub> at 0.12 µg/mL and MIC<sub>90</sub> at ≤0.25 µg/mL) displayed activity comparable to linezolid (>99.3% coverage at 2 µg/mL for all S. aureus; see Table 1).
- Among the 27 fusidic acid non-susceptible strains detected, 15 (55.5%) harbored acquired genes, including three strains (11.1% of the non-susceptible strains) positive for fusB and 12 strains (44.4%) positive for fusC.

- Mutations on fusA were detected in four strains [14.8% of the 27 (0.4%) resistant strains], all MRSA. These isolates carried previously described mutations M453I (one strain) L461S (one strain) and P404L combined with A71V (two strains from the same medical site).
- A 21-amino acid deletion on fusE was detected in two genetically similar (clonal) USA300-like strains from the same hospital. These strains demonstrated slow growth in culture media, suggesting a fitness cost for these small colony variants.

Table 2. Resistance mechanisms to fusidic acid detected in S. aureus isolated in the United States (2008-2009).

Year (no.)			No. by mechansism <sup>a</sup>						
	%MRSA	no. sites	fusA	fusB	fusC	fusE			
2008 (12)	33.3	7	M453I <sup>b</sup>	1	7	<b>2</b> <sup>c</sup>			
2009 (14)	50.0	12	L461S, A71V+P404L(2)d	2	5	-			
MRSA (11)	-	8	L461S, M453I, A71V+P404L(2) <sup>d</sup>	-	-	2 <sup>c</sup>			
MSSA (15)	-	11	-	3	12	-			

- Table 1. Antimicrobial activity of fusidic acid and comparator antimicrobial agents when tested against Staphylococcus aureus stratified by resistance patterns from USA medical centers (2008-2009).

Resistance pattern			_	CLSI <sup>b</sup>	EUCAST <sup>b</sup>	Resistance pattern			_	CLSI <sup>b</sup>	EUCAST <sup>b</sup>
Antimicrobial agent (no. strains) <sup>a</sup>	MIC <sub>50</sub>	MIC <sub>90</sub>	Range	%S / %R	%S / %R	Antimicrobial agent (no. strains) <sup>a</sup>	MIC <sub>50</sub>	MIC <sub>90</sub>	Range	%S / %R	%S / %R
Staphylococcus aureus (All; 7,340)						<u>ER + OX-R (751)</u>					
Fusidic acid	0.12	0.25	≤0.06 – 8	-/-	99.6 / 0.4	Fusidic acid	0.12	0.12	≤0.06 – 4	-/-	99.7 / 0.3
Linezolid	2	2	≤0.06 ->8	99.9 / 0.1	99.9 / 0.1	Linezolid	2	2	0.5 - 2	100.0 / 0.0	100.0 / 0.0
Tetracycline	≤2	≤2	≤2 ->8	95.5 / 3.8	94.5 / 5.5	Tetracycline	≤2	≤2	≤2 – 4	100.0 / 0.0	99.7 / 0.3
Trimethoprim/sulfamethoxazole	≤0.5	≤0.5	≤0.5 ->2	98.6 / 1.4	98.6 / 1.4	Trimethoprim/sulfamethoxazole	≤0.5	≤0.5	≤0.5 – 1	100.0 / 0.0	100.0 / 0.0
Daptomycin	0.25	0.5	≤0.06 – 4	99.9 / -	99.9 / 0.1	Daptomycin	0.25	0.5	0.12 - 1	100.0 / -	100.0 / 0.0
Tigecycline <sup>c</sup>	0.12	0.25	≤0.03 − 1	>99.9 / -	>99.9 / <0.1	Tigecycline <sup>c</sup>	0.12	0.25	≤0.03 – 0.5	100.0 / -	100.0 / 0.0
Vancomycin	1	1	≤0.12 – 2	100.0 / 0.0	100.0 / 0.0	Clindamycin	≤0.25	≤0.25	≤0.25 – 0.5	100.0 / 0.0	98.9 / 0.0
<u> 2008 (3,962)</u>						Quinupristin/dalfopristin	0.5	0.5	≤0.25 – 1	100.0 / 0.0	100.0 / 0.0
Fusidic acid	0.12	0.25	≤0.015 – 8	-/-	99.7 / 0.3	ER-R (729)					
Linezolid	2	2	0.25 ->8	99.9 / 0.1	99.9 / 0.1	Fusidic acid	0.12	0.25	≤0.06 – 8	-/-	99.7 / 0.3
Tetracycline	≤2	≤2	≤2 ->8	95.5 / 3.9	94.8 / 5.2	Linezolid	2	2	≤0.06 – 2	100.0 / 0.0	100.0 / 0.0
Trimethoprim/sulfamethoxazole	≤0.5	≤0.5	≤0.5 ->2	98.5 / 1.5	98.5 / 1.5	Tetracycline	≤2	≤2	≤2 – 4	100.0 / 0.0	99.6 / 0.4
Daptomycin	0.25	0.5	≤0.06 – 4	99.8 / -	99.8 / 0.2	Trimethoprim/sulfamethoxazole	≤0.5	≤0.5	≤0.5 – 2	100.0 / 0.0	100.0 / 0.0
Tigecycline <sup>c</sup>	0.12	0.25	≤0.03 – 1	>99.9 / -	>99.9 / <0.1	Daptomycin	0.25	0.5	0.12 – 1	100.0 / -	100.0 / 0.0
Vancomycin	1	1	≤0.12 – 2	100.0 / 0.0	100.0 / 0.0	Tigecycline <sup>c</sup>	0.12	0.25	≤0.03 – 0.5	100.0 / -	100.0 / 0.0
2009 (3,378)						Vancomycin	1	1	≤0.12 – 2	100.0 / 0.0	100.0 / 0.0
Fusidic acid	0.12	0.25	≤0.06 – 8	-/-	99.6 / 0.4	ER + CP-R (177)					
Linezolid	2	2	≤0.06 − >8	99.9 / 0.1	99.9 / 0.1	Fusidic acid	0.12	0.25	≤0.06 – 4	-/-	99.4 / 0.6
Tetracycline	<u>-</u> ≤2	<i>-</i> ≤2	≤2 ->8	95.4 / 3.7	94.2 / 5.8	Linezolid	2	2	0.5 – 2	100.0 / 0.0	100.0 / 0.0
Trimethoprim/sulfamethoxazole	<i>_</i> 2 ≤0.5	<i>_</i> 2 ≤0.5	≤0.5 - >2	98.6 / 1.4	98.6 / 1.4	Tetracycline	<i>≥</i>	≤2	≤2 – 4	100.0 / 0.0	99.4 / 0.6
Daptomycin	0.25	0.5	≤0.06 – 2	>99.9 / -	>99.9 / <0.1	Trimethoprim/sulfamethoxazole	= <u>∠</u> ≤0.5	<i>≟</i> 2 ≤0.5	=2 <del>-</del>	100.0 / 0.0	100.0 / 0.0
Tigecycline <sup>c</sup>	0.12	0.25	≤0.03 – 1	>99.9 / -	>99.9 / <0.1	Daptomycin	0.25	0.5	0.12 – 1	100.0 / -	100.0 / 0.0
Vancomycin	0.12	1	≤0.03 – 1 ≤0.12 – 2	100.0 / 0.0	100.0 / 0.0	Tigecycline <sup>c</sup>	0.23	0.3	0.12 − 1 ≤0.03 − 0.5	100.0 / -	100.0 / 0.0
MRSA (3,877)	Ī	Į.	30.12 - 2	100.07 0.0	100.0 / 0.0	Vancomycin	0.12	0.25	≥0.05 = 0.5 0.5 = 1	100.0 / 0.0	100.0 / 0.0
Fusidic acid	0.12	0.25	≤0.06 – 8	-/-	99.7 / 0.3	CL + CP + OX-R (137)	ı	Ī	0.5 – 1	100.07 0.0	100.07 0.0
Linezolid	2	2	<u>≤</u> 0.00 − 8 0.25 − >8	99.8 / 0.2	99.8 / 0.2	Fusidic acid	0.12	0.12	≤0.06 – 8	-/-	99.3 / 0.7
			0.25 - >6 ≤2 - >8	99.6 / 0.2	93.5 / 6.5	Linezolid			≤0.00 – 8 0.5 – 2	100.0 / 0.0	100.0 / 0.0
Tetracycline	≤2 <0.5	≤2 <0.5					2	2			
Trimethoprim/sulfamethoxazole	≤0.5	≤0.5	≤0.5 ->2	98.4 / 1.6	98.4 / 1.6	Tetracycline	≤2 10.5	≤2	≤2 – 4	100.0 / 0.0	99.3 / 0.7
Daptomycin	0.25	0.5	≤0.06 – 4	99.7 / -	99.7 / 0.3	Trimethoprim/sulfamethoxazole	≤0.5 0.25	≤0.5	≤0.5 – 2	100.0 / 0.0	100.0 / 0.0
Tigecycline <sup>c</sup>	0.12	0.25	≤0.03 – 1	99.9 / -	99.9 / 0.1	Daptomycin		0.5	0.25 – 1	100.0 / -	100.0 / 0.0
Vancomycin	1	1	0.25 – 2	100.0 / 0.0	100.0 / 0.0	Tigecycline <sup>c</sup>	0.12	0.25	0.06 - 0.5	100.0 / -	100.0 / 0.0
MSSA (3,463)						Vancomycin	1	1	0.5 - 2	100.0 / 0.0	100.0 / 0.0
Fusidic acid	0.12	0.25	≤0.06 – 8	-/-	99.6 / 0.4	ER + CL + CP-R (112)					
Linezolid	2	2	≤0.06 – 2	100.0 / 0.0	100.0 / 0.0	Fusidic acid	0.12	0.25	≤0.06 – 0.5	- / -	100.0 / 0.0
Tetracycline	≤2	≤2	≤2 ->8	96.1 / 2.9	95.6 / 4.4	Linezolid	2	2	0.5 - 2	100.0 / 0.0	100.0 / 0.0
Trimethoprim/sulfamethoxazole	≤0.5	≤0.5	≤0.5 ->2	98.7 / 1.3	98.7 / 1.3	Tetracycline	≤2	≤2	≤2	100.0 / 0.0	100.0 / 0.0
Daptomycin	0.25	0.5	≤0.06 – 1	100.0 / -	100.0 / 0.0	Trimethoprim/sulfamethoxazole	≤0.5	≤0.5	≤0.5 – 2	100.0 / 0.0	100.0 / 0.0
Tigecycline <sup>c</sup>	0.12	0.25	≤0.03 – 0.5	100.0 / -	100.0 / 0.0	Daptomycin	0.25	0.5	0.12 - 0.5	100.0 / -	100.0 / 0.0
Vancomycin	1	1	≤0.12 – 2	100.0 / 0.0	100.0 / 0.0	Tigecycline <sup>c</sup>	0.12	0.25	≤0.03 – 0.5	100.0 / -	100.0 / 0.0
ER + CP + OX-R (1,364)						Vancomycin	1	1	0.25 - 2	100.0 / 0.0	100.0 / 0.0
Fusidic acid	0.12	0.25	≤0.06 – 2	-/-	99.9 / 0.1	Pan-R (170)					
Linezolid	2	2	0.5 - 4	100.0 / 0.0	100.0 / 0.0	Fusidic acid	0.12	0.25	≤0.06 – 2	-/-	99.4 / 0.6
Tetracycline	≤2	≤2	≤2 – 4	100.0 / 0.0	99.2 / 0.8	Linezolid	1	2	0.25 ->8	97.1 / 2.9	97.1 / 2.9
Trimethoprim/sulfamethoxazole	≤0.5	≤0.5	≤0.5 – 2	100.0 / 0.0	100.0 / 0.0	Tetracycline	8	>8	≤2 ->8	45.9 / 48.2	42.9 / 57.1
Daptomycin	0.25	0.5	≤0.06 − 1	100.0 / -	100.0 / 0.0	Trimethoprim/sulfamethoxazole	≤0.5	>2	≤0.5 ->2	77.6 / 22.4	77.6 / 22.4
Tigecycline <sup>c</sup>	0.12	0.25	≤0.03 – 0.5	100.0 / -	100.0 / 0.0	Daptomycin	0.25	0.5	≤0.06 – 4	95.3 / -	95.3 / 4.7
Vancomycin	1	1	0.25 - 2	100.0 / 0.0	100.0 / 0.0	Tigecycline <sup>c</sup>	0.12	0.25	0.06 – 1	98.8 / -	98.8 / 1.2
ER + CL + CP + OX-R (1,156)						Vancomycin	1	1	0.25 - 2	100.0 / 0.0	100.0 / 0.0
Fusidic acid	0.12	0.25	≤0.06 – 4	-/-	99.6 / 0.4	a. Most prevalent resistance patterns no	ted among S.	aureus. MRS	SA = methicillin-resis	tant S. aureus; MSS	A = methicillin-
Linezolid	2	2	0.25 – 4	100.0 / 0.0	100.0 / 0.0	susceptible S. aureus. CL, clindamyc	n; CP, ciproflo	xacin; ER, er			
Tetracycline	≤2	<2	≤2 – 4	100 0 / 0 0	971/29	phenotype (resistance at least 5 antin			lactam cuccontibility	should be directed	by the evecillin t

100.0 / 0.0

100.0 / 0.0

100.0 / 0.0 100.0 / 0.0

- b. Criteria as published by the CLSI [2010] and EUCAST [2010], β-lactam susceptibility should be directed by the oxacillin test
- . USA-FDA breakpoints were applied [Tygacil Product Insert, 2005].

## Conclusions

- Fusidic acid non-susceptible strains (0.4% overall) showed varying resistance mechanisms. Plasmid-mediated resistance genes, such as fusB and fusC can be coselected by the use of antimicrobial agents other than fusidic acid.
- Although 99 to 100% of the resistant groups analyzed were susceptible to vancomycin, tigecycline, daptomycin, and linezolid according to CLSI interpretive criteria, fusidic acid was comparable or up to eight-fold more potent than these agents against these important multidrug-resistant (MDR) pathogens and is an agent that can be orally delivered.
- Fusidic acid exhibited potent activity against nearly all S. aureus (0.4% non-susceptible) similar to parenteral-only agents (daptomycin, tigecycline and vancomycin) and linezolid (modal MIC<sub>50</sub> at 0.12 μg/mL and MIC<sub>90</sub> at ≤0.25 μg/mL), emphasizing the usefulness of this antimicrobial agent to treat staphylococcal infections caused by MDR organisms in the USA.

#### References

- Biedenbach DJ, Rhomberg PR, Mendes RE, Jones RN (2010). Spectrum of activity. ation rates, synergistic interactions, and the effects of pH and serum proteins for fusidic acid (CEM-102). Diagn Microbiol Infect Dis 66: 301-307.
- 2. Castanheira M, Watters AA, Bell JM, Turnidge JD, Jones RN (2010). Fusidic acid resistance rates and prevalence of resistance mechanisms among Staphylococcus spp isolated in North America and Australia, 2007-2008. Antimicrob Agents Chemother 54:
- 3. Castanheira M, Watters AA, Mendes RE, Farrell DJ, Jones RN (2010). Occurrence and molecular characterization of fusidic acid resistance mechanisms among Staphylococcus
- spp. from European countries (2008). *J Antimicrob Chemother* 65: 1353-1358. 4. Clinical and Laboratory Standards Institute (2009). M07-A8. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically; approved standard: eighth edition. Wayne, PA: CLSI.
- 5. Clinical and Laboratory Standards Institute (2010). *M100-S20. Performance standards for* antimicrobial susceptibility testing: 20th informational supplement. Wayne, PA: CLSI.
- 6. Howden BP, Grayson ML (2006). Dumb and dumber--the potential waste of a useful antistaphylococcal agent: emerging fusidic acid resistance in Staphylococcus aureus. Clin
- Infect Dis 42: 394-400. 7. Jones RN, Biedenbach DJ, Roblin PM, Kohlhoff SA, Hammerschlag MR (2010). Update of fusidic acid (CEM-102) tested against Neisseria gonorrhoeae and Chlamydia trachomatis. Antimicrob Agents Chemother 54: 4518-4519.
- 8. Jones RN, Castanheira M, Rhomberg PR, Woosley LN, Pfaller MA (2010). Performance of Fusidic Acid (CEM-102) susceptibility testing reagents: Broth microdilution, disk diffusion, and Etest methods as applied to Staphylococcus aureus. J Clin Microbiol 48: 972-976.
- 9. Jones RN, Ross JE (2009). Initial quality control (QC) ranges for CEM-102 (fusidic acid [FA]) using the CLSI multi-laboratory M23-A3 study design 49th ICAAC, September 12-15, 2009, San Francisco, CA, USA.
- 10. Pfaller MA, Castanheira M, Sader HS, Jones RN (2010). Evaluation of the activity of fusidic acid tested against contemporary Gram-positive clinical isolates from the USA and Canada. Int J Antimicrob Agents 35: 282-287.
- 11. Rhomberg PR, Mendes RE, Becker HK, Fedler KA, Sader HS, Jones RN (2009). Update on the spectrum of CEM-102 (fusidic acid [FA]) against contemporary wildtype (WT) bacterial species including mutational resistance (R) analysis, and synergy testing. Abstr. 203. 47th IDSA, October 29-November 1, 2009, Philadelphia, Pennsylvania, USA.
- 12. Rhomberg PR, Woosley LN, Sader HS, Jones RN (2009). Contemporary antimicrobial activity of CEM-102 (fusidic acid [FA]) against Canadian isolates of staphylococci and streptococci (2001-2006). Abstr. 202. 47th IDSA, October 29-November 1. 2009 Philadelphia, Pennsylvania, USA.
- 13. Rhomberg PR, Woosley LN, Sader HS, Jones RN (2009). Performance of CEM-102 (fusidic acid [FA]) susceptibility testing reagents; broth microdilution, disk diffusion and Etest methods. Abstr. 261. 47th IDSA, October 29-November 1, 2009, Philadelphia, Pennsylvania, USA.
- 14. Turnidge J, Collignon P (1999). Resistance to fusidic acid. *Int J Antimicrob Agents* 12 Suppl