IDWEEK 2024 | Poster #P1361

Cefiderocol Activity against Pseudomonas aeruginosa, Including **Resistant Subsets and Isolates Carrying Carbapenemase** β-lactamase Genes, from United States Hospitals (2020–2023)

RE Mendes, JM Maher, A Scullin, M Karr, JH Kimbrough, M Castanheira Element Iowa City (JMI Laboratories), North Liberty, IA, USA

Introduction

- Pseudomonas aeruginosa possess various intrinsic treatment-limiting resistance mechanisms, leading to decreased antibiotic permeability.
- Isolates may acquire β -lactamase genes, such as those encoding class A carbapenemases and especially class B metallo- β -lactamases further decreasing susceptibility to numerous β -lactams.
- Cefiderocol is approved by the US Food and Drug Administration (FDA) for the treatment of complicated urinary tract infections, including pyelonephritis, as well as hospital-acquired bacterial pneumonia and ventilator-associated bacterial pneumonia.
- Cefiderocol is a siderophore cephalosporin with broad activity against Gramnegative bacteria, including multidrug-resistant (MDR) organisms like carbapenemresistant Enterobacterales (CRE), carbapenem-resistant *Pseudomonas aeruginosa*, and Acinetobacter baumannii.
- In this study, the activities of cefiderocol and comparator agents were evaluated against *P. aeruginosa* causing infections in US hospitals, including resistant subsets, as part of the SENTRY Antimicrobial Surveillance Program during 2020–2023.

Materials and Methods

Bacterial organisms

- This study comprised a collection of 4,400 *P. aeruginosa* cultured from various clinical specimens in patients hospitalized in 38 medical centers in all 9 US Census Divisions during 2020–2023. Only consecutive isolates (1 per patient infection episode) responsible for documented infections according to local criteria were included.
- Bacterial identification was confirmed by standard algorithms supported by matrixassisted laser desorption ionization-time of flight mass spectrometry (Bruker Daltonics, Bremen, Germany).

Susceptibility testing

- Isolates were tested for susceptibility by broth microdilution following the Clinical and Laboratory Standards Institute (CLSI) M07 (2018) guidelines.
- Frozen-form broth microdilution panels were manufactured by Element Iowa City (JMI Laboratories) (North Liberty, IA, USA) and contained cation-adjusted Mueller-Hinton broth for comparator agents.
- Susceptibility testing for cefiderocol used broth microdilution panels containing iron-depleted media per CLSI guidelines.
- Quality assurance was performed by sterility checks, colony counts, and testing CLSI-recommended quality control reference strains.
- Cefiderocol MIC results were interpreted according to the CLSI and FDA criteria, whereas comparator agent MIC values were interpreted based on CLSI breakpoints.
- Carbapenem-nonsusceptible isolates were those nonsusceptible to imipenem and/or meropenem based on CLSI criteria (MIC, ≥4 mg/L). MDR was classified as nonsusceptible to ≥3 drug classes using CLSI breakpoints; extensively drug-resistant (XDR) was defined as nonsusceptible to all but 2 or fewer drug classes using CLSI breakpoints.

Screening of β -lactamase genes

- Laboratories.
- known β -lactamase genes.

Results

- P. aeruginosa.

- (Table 1).

Selected isolates had total genomic DNA extracted by the fully automated Thermo Scientific[™] KingFisher[™] Flex Magnetic Particle Processor (Cleveland, OH, USA), which was used as input material for library construction.

DNA libraries were prepared using the Nextera[™] or Illumina DNA Prep[™] library construction protocol (Illumina, San Diego, CA, USA) following the manufacturer's instructions and were sequenced on MiSeq or NextSeq Sequencer platforms at JMI

FASTQ format sequencing files for each sample set were assembled independently using *de novo* assembler SPAdes 3.15.3. An in-house software was applied to align the assembled sequences against a comprehensive in-house database containing

• A total of 14.6% (643/4,400) and 4.7% (206/4,400) *P. aeruginosa* isolates were categorized as MDR and XDR, respectively (Table 1).

In addition, 22.3% (980/4,400) *P. aeruginosa* were carbapenemnonsusceptible, and 99.1% (971/980) were carbapenemase-negative.

Most carbapenem-nonsusceptible *P. aeruginosa* originated from pneumonia patients, whereas smaller percentages were from skin and skin-structure infections (16%), bloodstream infections (8%), and urinary tract infections (7%) (Figure 1).

• Cefiderocol and β -lactam/ β -lactamase inhibitor (BL/BLI) combinations showed susceptibilities of >96% against all *P. aeruginosa*, except for piperacillin-tazobactam (79.9% susceptible) (Table 1).

• Cefiderocol (90.8–99.4% susceptible) showed MIC₅₀ of 0.12 mg/L and MIC₉₀ of 0.5–1 mg/L against MDR, XDR, and carbapenem-nonsusceptible isolates (Table 1).

BL/BLI combinations showed various lower degrees of susceptibilities (2.9–84.1% susceptible) against the MDR and XDR subsets.

Imipenem-relebactam, ceftazidime-avibactam, and ceftolozane-tazobactam showed susceptibilities of 86.9–90.2% against carbapenem-nonsusceptible

Piperacillin-tazobactam inhibited 49.7% of carbapenem-nonsusceptible isolates at the CLSI breakpoint for susceptibility.

Cefiderocol (100% susceptible) inhibited all *P. aeruginosa* carrying carbapenemases at $\leq 2 \text{ mg/L}$ (Table 1).

Other comparators showed susceptibilities of <34% against *P. aeruginosa* carrying carbapenemases.

 Cefiderocol (MIC_{50/90}, 0.12/0.5 mg/L; 96.8–99.4% susceptible), imipenemrelebactam (MIC_{50/90}, 1/2 mg/L; 90.0% susceptible), and ceftolozane-tazobactam (MIC_{50/90}, 1/4 mg/L; 90.9% susceptible) were the most active agents against carbapenem-nonsusceptible *P. aeruginosa* without carbapenemase genes

Ceftazidime-avibactam (87.4% susceptible) showed a marginal and lower activity against this subset.

Figure 1. Distribution of infection types^a caused by carbapenem-nonsusceptible *P. aeruginosa*

Table 1. Activity of cefiderocol and β-lactam-β-lactamase inhibitor combinations against *P. aeruginosa* and resistant subsets from the USA

Phenotype/genotype ^a (No. tested)	MIC ₅₀ /MIC ₉₀ in mg/L (% susceptible by CLSI/FDA criteria) ^b					
	FDC	IMR	CZA	C/T	P/T	ME
All (4,400)	0.12/0.25 (99.9/98.5)	0.25/1 (97.6)	2/4 (96.7)	0.5/2 (97.4)	4/128 (79.9)	0.5/8 (8
MDR (643)	0.12/0.5 (99.1/95.0)	1/4 (84.1)	8/16 (78.1)	2/16 (83.8)	128/>128 (11.4)	8/32 (1
XDR (206)	0.12/1 (98.5/90.8)	2/8 (56.3)	8/>32 (62.1)	4/>16 (69.9)	128/>128 (2.9)	16/32 (
Carbapenem-nonS (980)	0.12/0.5 (99.4/96.6)	1/4 (89.5)	4/16 (86.9)	1/4 (90.2)	32/>128 (49.7)	8/32 (1
Carbapenemase-positive ^c (9)	0.12/— (100/77.8)	>8/— (33.3)	16/— (33.3)	>16/— (11.1)	128/— (22.2)	>32/—
Carbapenemase-negative (971)	0.12/0.5 (99.4/96.8)	1/2 (90.0)	4/16 (87.4)	1/4 (90.9)	16/>128 (50.0)	8/32 (1

bbreviations: FDC. cefiderocol: IMR. imipenem-relebactam: CZA. ceftazidime-avibactam; C/T. ceftolozane-tazobactam; P/T. piperacillin-tazobactam; MER, meropenen ^a Carbapenem-nonS, isolates nonsusceptible to imipenem and/or meropenem based on CLSI criteria (MIC values ≥4 mg/L); MDR, multidrug-resistant isolates classified as nonsusceptible to ≥3 drug classes using CLSI breakpoints; XDR, extensively drug-resistant, defined as nonsusceptible to all but 2 or fewer drug classes using CLSI breakpoints. ^b Cefiderocol MIC results were interpreted according to the CLSI/FDA criteria, whereas comparator agent MIC values were interpreted based on CLSI criteria. ^c Includes $bla_{GES-5}(1)$, $bla_{IMP-1}(1)$, $bla_{IMP-13}(1)$, $bla_{KPC-2}(1)$, $bla_{NDM-1}(2)$, and $bla_{VIM-2}(3)$.



^a BSI, bloodstream infections; SSSI, skin and skin structure infections; and UTI, urinary tract infections.

Conclusions

- Cefiderocol showed potent activity against *P. aeruginosa* clinical isolates from US hospitals, including resistant subsets, and carbapenem-nonsusceptible isolates with or without carbapenemase genes.
- These data demonstrated cefiderocol in vitro activity against P. aeruginosa resistant subsets, for which antibiotic treatment options are limited.

Acknowledgements

This research and poster presentation were sponsored by Shionogi & Co., LTD.

References

1. Clinical and Laboratory Standards Institute. 2018. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically. M07 11th Edition. Wayne, PA,

2. Clinical and Laboratory Standards Institute. 2024. Performance standards for antimicrobial susceptibility testing. M100 34th Edition. Wayne, PA, USA.

3. FDA Susceptibility Test Interpretive Criteria: https://www.fda.gov/drugs/development -resources/antibacterial-susceptibility-test-interpretive-criteria. Accessed April 2024.

4. Karlowsky JA, Hackel MA, Takemura M, Yamano Y, Echols R, Sahm DF. 2022. In vitro susceptibility of Gram-negative pathogens to cefiderocol in five consecutive annual multinational SIDERO-WT Surveillance Studies, 2014 to 2019. Antimicrob Agents Chemother. 66: e0199021.

5. Mendes RE, Jones RN, Woosley LN, Cattoir V, Castanheira M. 2019. Application of next-generation sequencing for characterization of surveillance and clinical trial isolates: Analysis of the distribution of β -lactamase resistance genes and lineage background in the United States. Open Forum Infect Dis 6: S69–S78.

6. Ong'uti S, Czech M, Robilotti E, Holubar M. 2022. Cefiderocol: A new cephalosporin stratagem against multidrug resistant Gram-negative bacteria. Clin Infect Dis. 74: 1303–1312.

Contact



Rodrigo E. Mendes, Ph.D. JMI Laboratories 345 Beaver Kreek Centre, Suite A North Liberty, Iowa 52317 Phone: (319) 665-3370 Fax: (319) 665-3371 Email: rodrigo.mendes@element.com



To obtain a PDF of this poster: Scan the QR code or visit https://www

.jmilabs.com/data/posters/IDWeek2024 _23-SHI-09_A5_PSA_IDWeek2024.pdf

Charges may apply. No personal information is stored.

```
7.9)
```

```
5.9)
```